

# Measuring Double-Stranded DNA Concentration Using the Quantus™ Fluorometer with the Quant-iT™ PicoGreen® dsDNA Assay Kit

Promega Corporation



## Materials Required

- Quantus™ Fluorometer (Cat.# E6150)
- 0.5ml PCR Tubes (Axygen Cat.# PCR-05-C, available through Fisher or VWR)
- Quant-iT™ PicoGreen® dsDNA Assay Kit (Life Technologies Cat.# P7589)

**Caution:** We recommend the use of gloves, lab coats and eye protection when working with these or any chemical reagents.

**Protocol:** *Quantus™ Fluorometer Operating Manual #TM396* is available at: [www.promega.com/protocols/](http://www.promega.com/protocols/)

Detecting and quantitating small amounts of DNA are important steps in a wide variety of biological applications. These include standard molecular biology techniques such as constructing cDNA libraries and purifying DNA fragments for subcloning, as well as diagnostic techniques such as quantitating DNA amplification products and detecting DNA molecules in drug preparations.

The most commonly used technique to determine nucleic acid concentration is measuring absorbance at 260nm ( $A_{260}$ ). The major disadvantages of the absorbance-based method include: the inability to distinguish among DNA (both single- and double-stranded), RNA and nucleotides, interference caused by contaminants commonly found in nucleic acid preparations, and the relative insensitivity of the assay (traditional spectrophotometric assays cannot determine nucleic acid concentrations below 2 $\mu$ g/ml). The use of sensitive, fluorescent nucleic acid stains alleviates many of these problems.

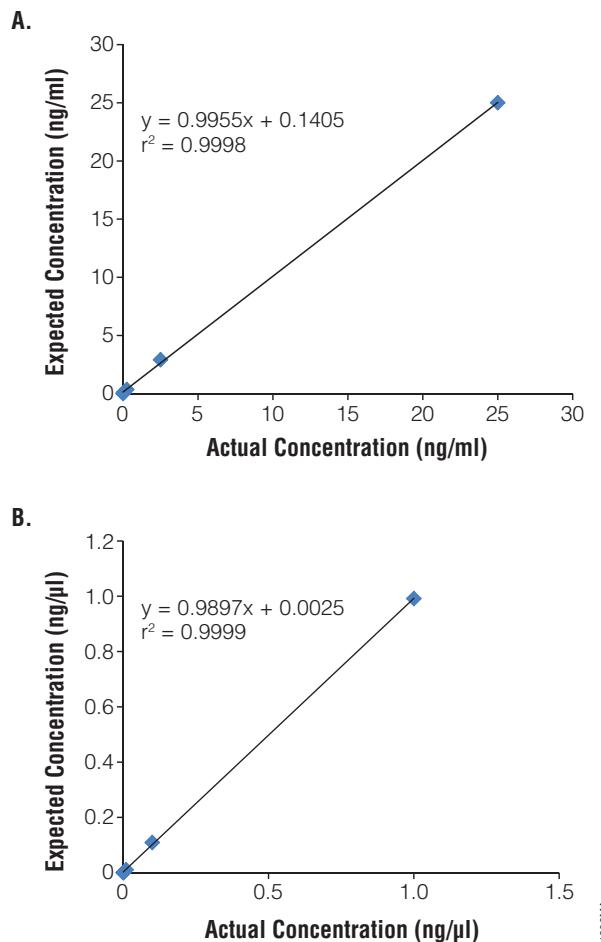
The Quant-iT™ PicoGreen® dsDNA Assay Kit can be used with the Quantus™ Fluorometer. The assay quantitates double-stranded DNA (dsDNA) in solution at a final assay concentration of 1ng/ $\mu$ l–250pg/ml of dsDNA.

This Application Note describes the protocol for using the Quant-iT™ PicoGreen® dsDNA Assay Kit with the Quantus™ Fluorometer.

## Protocol

1. Create a custom protocol on the Quantus™ Fluorometer by selecting “New” from the menu list on the Protocol screen, and name the protocol by using the up or down buttons. For the high-concentration standard, enter the standard value of 1ng/ $\mu$ l. For the low-concentration standard, enter the standard value of 25ng/ml. Select the Blue channel, and save the protocol.
2. Equilibrate all reagents to room temperature.
3. Dilute the Quant-iT™ PicoGreen® reagent 1:200 in 1X TE buffer to make a reagent working solution. For example, add 10 $\mu$ l of Quant-iT™ PicoGreen® reagent to 1,990 $\mu$ l of 1X TE buffer, and mix.

4. Add 100 $\mu$ l of the Quant-iT™ PicoGreen® reagent working solution to a 0.5ml PCR tube containing 100 $\mu$ l of 1X TE buffer. This will be the blank sample used in Step 8. Protect from light.
  5. Prepare the dsDNA standard. For a high-concentration standard protocol, dilute the dsDNA standard (Component C) to 2ng/ $\mu$ l by adding 40 $\mu$ l of Component C to 1,960 $\mu$ l of 1X TE buffer, and mix. For a low-concentration standard protocol, dilute the dsDNA standard to 50ng/ml by adding 1 $\mu$ l of Component C to 1,999 $\mu$ l of 1X TE buffer, and mix. Add 100 $\mu$ l of standard to 100 $\mu$ l of Quant-iT™ PicoGreen® reagent working solution in a 0.5ml PCR tube, and mix. This will be the standard sample used in Step 8. Protect from light.
  6. Add 100 $\mu$ l of unknown sample and 100 $\mu$ l of Quant-iT™ PicoGreen® reagent working solution to a 0.5ml PCR tube, and mix.
- Note:** If the volume of the unknown DNA sample is less than 100 $\mu$ l, add 1X TE buffer to bring to a final volume of 100 $\mu$ l. For example, dilute a 1 $\mu$ l sample with 99 $\mu$ l of 1X TE buffer, then add 100 $\mu$ l of reagent working solution for a total volume of 200 $\mu$ l.
7. Incubate the blank, standard and unknown samples at room temperature for 5 minutes, protected from light.
  8. Select the custom protocol created in Step 1. Go to the Calibration screen and read the blank and standard samples prepared in Steps 4 and 5. Save the calibration.
  9. Enter the volume of the unknown sample and desired concentration units.
- Note:** This volume is the amount of sample that is added for the quantitation. For example, if 1 $\mu$ l of sample was mixed with 99 $\mu$ l of 1X TE buffer then added to 100 $\mu$ l of reagent working solution for a total volume of 200 $\mu$ l in the tube, then the volume entered on this screen is 1 $\mu$ l.
10. Place the unknown sample into the tube holder, and close the lid. The instrument will automatically measure fluorescence when the lid is closed, and the calculated nucleic acid concentration will be displayed.



**Figure 1. Measuring dsDNA concentration using the Quant-iT™ PicoGreen® dsDNA reagent and Quantus™ Fluorometer.** Standard curves were generated per manufacturer's instructions to demonstrate the linearity of the Quantus™ Fluorometer. Samples were run in duplicate. **Panel A.** Assay linearity using the low-concentration standard curve. **Panel B.** Assay linearity using the high-concentration standard curve.

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